

**IN-VITRO THROMBOLYTIC AND ANTICOAGULANT ACTIVITY
OF ETHANOLIC EXTRACT OF *Carum copticum* AND *Triconella
foenum-graecum***

A Thesis

By

MD. AKTER HOSSEN
Registration No. 1305106
Semester: July-December, 2014
Session: 2013-14

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**MASTER OF SCIENCE (M.S.)
IN
PHARMACOLOGY**

**DEPARTMENT OF PHYSIOLOGY AND PHARMACOLOGY
HAJEE MOHAMMAD DANESH SCIENCE AND TECHNOLOGY
UNIVERSITY, DINAJPUR-5200**

DECEMBER, 2014

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*Submitted to the
Department of Physiology & Pharmacology
Hajee Mohammad Danesh Science and Technology University, Dinajpur,
In Partial fulfillment of the requirements
For the degree of*

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Approved as to the style and content by



(Dr. Md Bazlar Rashid)
Supervisor



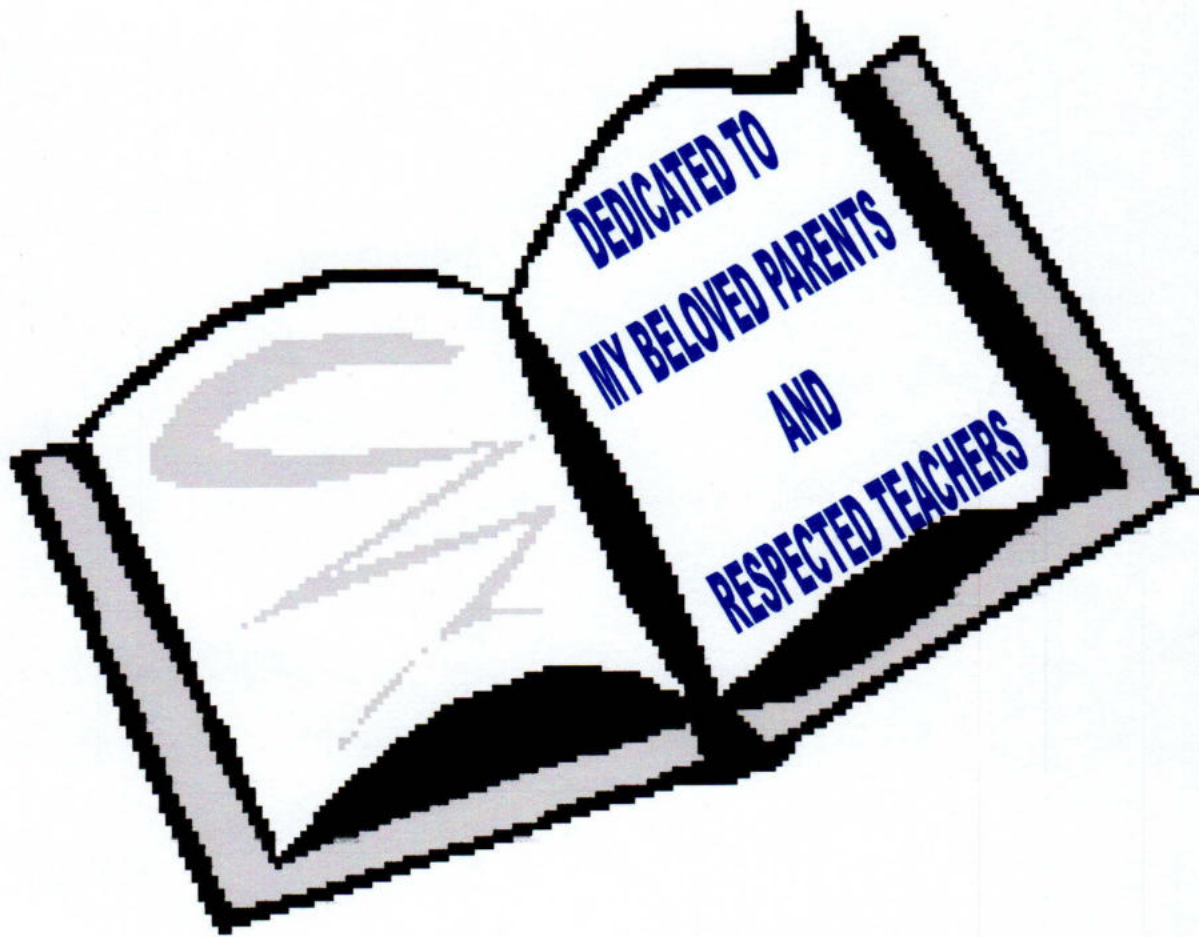
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The Author

December 2014

ABSTRACT

An experiment was carried out in the laboratory of the department of physiology and pharmacology, Hajee Mohammad Danesh Science and Technology University, Dinajpur, during the period of July-December/2014. Some plant and spices were collected from Dinajpur district. Primarily ethanol extract was prepared from those plant and spices. 20 micropipette was taken filled with 0.5 ml human blood and 3 micropipette was taken as excess micropipette for control. These two kind of micropipette were screened for assessment their comparative efficacy of clotted blood lysis. Ethanol extract was used at a dose of 100 micro ml/ micropipette. Then this micropipette was incubated at 37 degree centigrade temperature for 1hr 45 minutes. After incubation I found 4 plant and 2 spices has anticoagulant property. These plant are Fern, Arum, Cannabis Tezpata and spices are Ajawin and Pesta. Finally I was experimented with Ajawin and Methi *in-vitro* thrombolytic and anticoagulant property. Anticoagulant and thrombolytic property was experimented in laboratory compared with Streptokinase (SK). Streptokinase showed 100% efficacy after treatment. Efficacy was recorded by compare among streptokinase control treated extract. Ethanol ajawin extract lysis clotted blood 57.33%.

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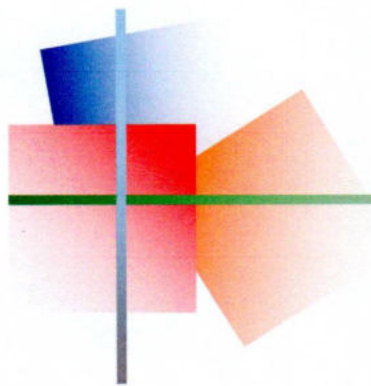
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LIST OF ABBREVIATIONS

B.wt.	:	Body weight
BAU	:	Bangladesh Agricultural University
Conc.	:	Concentration
Cu mm	:	Cubic millimeter
d.w.	:	Drinking water
ESR	:	Erythrocyte Sedimentation Rate
<i>et al.</i>	:	Associates
Fig.	:	Figure
Gm	:	Gram
Hb	:	Hemoglobin
i.e.	:	That is
J.	:	Journal
Kg	:	Kilogram
Lit	:	Liter
Ltd.	:	Limited
Mg	:	Milligram
ml	:	Milliliter
mm ³	:	cubic millimeter
No.	:	Number
PBS	:	Phosphate Buffer Solution
PCV	:	Packed Cell
PM	:	Population Mean
SE	:	Standard Error

LIST OF ABBREVIATIONS (contd.)

SM	:	Sample Mean
TEC	:	Total Erythrocyte Count
Vol.	:	Volume
μg	:	Microgram
%	:	Percent
&	:	And
@	:	At the rate of
<	:	Less than
>	:	Greater than
\pm	:	Plus minus
0°C	:	Degree centigrade



Chapter 1
INTRODUCTION

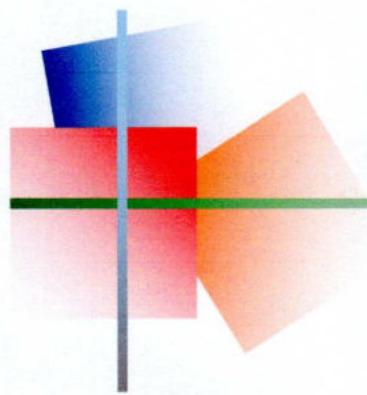
CHAPTER 1

INTRODUCTION

Man has many problem for their life. Man has many diseases such as cardiovascular disease, urogenital disease, nutritional disease etc. When man affected with cardiovascular or myocardial or cerebral infections. Myocardial or cerebral infractions are the serious consequences in Atherothrombotic diseases leading to death and the side effects produced by consecutive use of thrombolytic agent like t-PA, Urokinase and streptokinase to treat these diseases has become a global concern. Thrombolytic agents such as tissue plasminogen activator, urokinase, streptokinase (SK), etc are used to dissolve the already formed clots in the blood vessels . However, these drugs have certain limitations which cause serious and sometimes fatal consequences including hemorrhage, severe anaphylactic reaction, lacked specificity, etc. Moreover, as a result of immunogenicity multiple treatments with SK in a given patient are restricted. Methi and ajawin has anticoagulant and thrombolytic property respectively. The present study was carried out to evaluate the thrombolytic and anticoagulant activity of ethanolic extract of ajawin methi spices by in vitro method. Extraction was carried out using Soxhlet apparatus. The ajawin ethanol extract showed highest thrombolysis than methi extract. But methi showed highest anticoagulant property than ajawin. Concentration of phytochemicals and incubation time were directly proportional to the clot lysis. Application of the present study may be accessible for greater section of the society for treatment of cardiovascular diseases. Now-a-day synthetic drug like streptokinase so expensive has many side effect. As Methi and Ajawin has thrombolytic anticoagulant activity so the extraction of Methi and ajawin used as treatment for cardiovascular disease against t-PA, Urokinase and streptokinase. Which will be cheapest than t-PA, Urokinase and streptokinase.

In concern to that consideration my study was designed for the following objectives

- To observe the performance of thrombolytic activity of spices.
- To observe the performance of anticoagulant activity of spices.



Chapter 2

REVIEW OF LITERATURE

CHAPTER 2

REVIEW OF LITERATURE

Cardiovascular disease serious human diseases, causing millions of deaths every year. Thrombosis is one of the leading cause of thromboembolic disorders affecting million persons worldwide. Several plants used for the treatment of thromboembolic diseases in different systems of traditional medicine have shown anticoagulant/antithrombotic activity and such plants claimed in the traditional system still remain to be scientifically investigated. For more than five decades, anticoagulant drugs consisting of heparins, vitamin K antagonists, and their derivatives have been the major players in the clinical setting. Although their efficacy remains undisputed, the deleterious life-threatening side effects of these drugs have also been well documented. Plants may serve as the alternative sources for the development of new anticoagulant agents due to their biological activities. There is compelling scientific evidences demonstrating that the consumption of dietary anticoagulants or phytochemicals with anticoagulant properties can ultimately reduce or eliminate the risks of thromboembolic diseases. Prothrombin time (PT) is measure of the extrinsic coagulation pathway.

Mohammad Shahadat Hossain, et al. (2012) In-Vitro Thrombolytic and Anti-inflammatory Activity of *Swertia Chirata*. Ethanolic extract of *Swertia chirata* was assessed for its thrombolytic, anti-inflammatory activity and phytochemical screening. *In vitro* anti-inflammatory activity was evaluated using albumin denaturation. Aspirin was used as a standard drug for the study of anti-inflammatory activity. The ethanol extract of *Swertia chirata* showed mean inhibition of protein denaturation 45.31 ± 0.000576 whereas, for control group it was found to be 50.00 ± 0.00177 . In thrombolytic activity using *in vitro* clot lysis assay method, the crude ethanol extract was found to have significant, thrombolytic test showed a maximum effect of 40.38% while the standard streptokinase showed 69.35ct.

Narjis Hadi Mansoor (2013) In vitro study of the anticoagulant activity of some plant extracts examined by prothombin time, anticoagulant activity, red onion, garlic oil, grape oil.

Parvin, et al. (2013) Phytochemical screenings, thrombolytic activity and antimicrobial properties of the leaf extracts of *Lablab purpureus*. In this present study, the leaves extracts of *Lablab purpureus* were subjected to the thrombolytic activities were assessed by using human erythrocyte and the results were compared with standard streptokinase (SK). On the other hand, leaves extracts of *L. purpureus* revealed moderate antibacterial activity against some microorganisms used in the screening. Preliminary phytochemical investigation suggested the presence of reducing sugar group, tannins, saponins and alkaloids.

Y., Sai Sandeep, et al. (2012) Evaluation of in vitro thrombolytic activity of phytochemicals in *Bacopa monnieri* Linn. Myocardial or cerebral infarctions are the serious consequences in Atherothrombotic diseases leading to death and the side effects produced by consecutive use of thrombolytic agent like t-PA, Urokinase and streptokinase to treat these diseases has become a global concern. *Bacopa monnieri* has been used for centuries in Ayurveda system of medicine as a memory vitalizer and as a liver and heart tonic. The present study was carried out to evaluate the thrombolytic activity of ethanolic, methanolic, acetone and aqueous extract of different parts (root, stem and leaf) of *B. monnieri* by in vitro method. Extraction was carried out using Soxhlet apparatus. The leaf ethanolic extract showed highest thrombolysis followed by aqueous, methanol and acetone extract. Concentration of phytochemicals and incubation time were directly proportional to the clot lysis. Application of the present study may be accessible for greater section of the society for treatment of cardiovascular diseases.

Md. R. Al-Mamun et al. (2012), Thrombolytic activity of some spices and plants available in Bangladesh. Thrombolytic activities of some plants, namely *Tamarindus indica* (Fabaceae), *Flemingia congesta* (Fabaceae), *Lawsonia inermis*

(Lythraceae), *Mesua nagassarium* (Clusiaceae, and spices, namely *Coriandrum sativum* (Apiaceae), *Curcuma longa* (Zingiberaceae), *Cinnamomum tamala* (Lauraceae), *Nigella sativa* (Ranunculaceae), *Eugenia aromaticum* (Myrtaceae), available in Bangladesh, were evaluated using an *in vitro* model. The thrombolytic activity in terms of percentage of weight loss of *in vitro* formed clots were found as *C. sativum* $43.25 \pm 7.18\%$, *C. longa* $53.32 \pm 4.96\%$, *C. tamala* $22.10 \pm 3.18\%$, *N. sativa* $28.49 \pm 3.72\%$, *E. aromaticum* $32.18 \pm 3.10\%$, *T. indica* $28.91 \pm 2.29\%$, *F. congesta* $35.27 \pm 7.35\%$, *L. inermis* $62.40 \pm 5.04\%$, *M. nagassarium* bark $39.54 \pm 7.15\%$ and *M. nagassarium* leaf $46.75 \pm 3.97\%$ with reference to the negative control distilled water $8.37 \pm 1.18\%$ and positive control streptokinase $84.63 \pm 1.03\%$. Through our study, it was found that *L. inermis* and *C. longa* possess thrombolytic property that could lyse blood clots *in vitro*.

Md. Al Amin Sikder, et al. (2011) Journal of Pharmacy and Nutrition Sciences examined evaluation of thrombolytic activity of four Bangladeshi medicinal plants, as a possible renewables for thrombolytic Compounds. Four Bangladeshi medicinal plants *Sansevieria trifasciata*, *Justica gendarussa*, *Hydnocarpus kurzii* and *Mesua nagassarium* have been investigated for their *in vitro* thrombolytic activity. The clot lysis activity was assessed by addition of the test material to the pre-clotted blood and incubation for 90 min. at 37°C and was expressed as % lysis of clot. Each of the plant was extracted with methanol at room temperature and the concentrated methanolic extract was fractionated by the modified Kupchan partitioning method to provide pet-ether, carbon tetrachloride, chloroform and aqueous soluble fractions. Among the four plants the aqueous soluble fraction of *M. nagassarium*, carbon tetrachloride soluble fraction of *H. Kurzii*, aqueous soluble fraction of methanolic extract of *S. trifasciata* exhibited highest thrombolytic activity with clot lysis value of 50.86%, 47.50%, and 47.10% respectively. However, the pet ether and carbon tetrachloride soluble fraction of methanolic extract of *J. gendarussa* demonstrated significant thrombolytic activity as evident from 45.93% and 45.47% lysis of clot,

respectively. Standard streptokinase was used as positive control which exhibited 61.50% lysis of clot while the negative control water revealed 2.56% lysis of clot.

J.H. Evangelista *et al.* (2012) examined Preliminary Assessment of In vitro Anticoagulant Activity vs. Heparin 1,000I.U. and Cytotoxicity of Selected Philippine Medicinal Plants. One main cause of mortality in developing countries is thromboembolic disorders such as pulmonary emboli, deep vein thrombosis and heart attacks. Several agents and interventions were available; however, there are still side effects that are acquired through these therapies. Herbal plants are popularly used nowadays in drug discovery due to their ancient medicinal use. Philippines, a tropical country, have a variety of herbal plants. The aim of the study was to assess the anticoagulant activity and cytotoxicity of both flesh and peels/seeds of selected plants available in the Philippines. The plants *Allium sativum*, *Cucurma longa*, *Ananas comosus* and *Lycopersicum esculentum* were used in the in vitro method using Heparin and water as controls. Percentage clot lysis of the plants were 18.30%, 21.77%, 21.85%, 35.91%, 15.67% and 24.52%, respectively. Only *C. longa* peel extracts and *L. esculentum* showed clot lysis beyond the negative control. *C. longa* peels showed a higher percentage clot lysis as compared with Heparin. Using one-way ANOVA, statistics showed the p value of 0.674574 from the clot lysis activity of all plant extracts. Cytotoxicity of the herbal plants was also determined using brine shrimp lethality assay (BSLA). The LC50 values of the herbal extracts ranges from 6.72 to 31.2µg/mL. *A. sativum* and *C. longa* flesh have the lowest values (most potent among the extracts). Descriptively, *C. longa* peels extract exhibited promising clot lysis activity, however further studies are still needed to strengthen the effectiveness of these Philippine plants as anticoagulant. Furthermore, it was also seen in this study that fruit and vegetable peels have prospective therapeutic application. It will not only benefit the industrial waste products reduction, but it can also serve as an alternative source of pharmacologic agents.

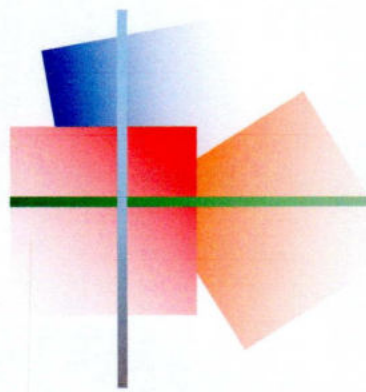
Jannat-e-Zereen and Gwyneth Ingram (2012) A Possible Involvement of *ACR4*, a Receptor Like Kinase, in Plant Defence Mechanism, *Bangladesh Pharmaceutical Journal*.

M Atiar Rahman, Rabeya Sultana, Talha Bin Emran, M Saiful Islam, M Ashiqur Rahman, Joti Sankhar Chakma, Harun-ur Rashid and Chowdhury Mohammad Monirul Hasan (2013) Effects of organic extracts of six Bangladeshi plants on *in vitro* thrombolysis and cytotoxicity, *BMC Complementary and Alternative Medicine*.

Md. Rakib Al-Mamun^{1,2}, Nabiha Amrin², Jahura Begum² and Md. Abdul Mazid¹ (2012) Thrombolytic activity of some spices and plants available in Bangladesh.

Sweta Prasad, Rajpal Singh Kashyap, and Hatim F Daginawala (2012) Effect of *Fagonia Arabica* (Dhamasa) on *in vitro* thrombolysis, BioMed Central. Atherothrombotic diseases such as myocardial or cerebral infarction are serious consequences of the thrombus formed in blood vessels. Thrombolytic agents are used to dissolve the already formed clots in the blood vessels; however, these drugs have certain limitations which cause serious and sometimes fatal consequences. Herbal preparations have been used since ancient times for the treatment of several diseases. Herbs and their components possessing antithrombotic activity have been reported before; however, herbs that could be used for thrombolysis has not been reported so far. This study's aim was to investigate whether herbal preparations (aqueous extract) possess thrombolytic activity or not.

Y., Sai Sandeep; Panigrahi, Mamata; C., Divya. G.; B., Beena D. (2011) Evaluation of *in vitro* thrombolytic activity of phytochemicals in *Bacopa monnieri* Linn, January 2012, *Journal of Pharmacy Research*; Jan2012, Vol. 5 Issue 1, p100, Academic Journal.



Chapter 3

MATERIALS AND METHODS

CHAPTER 3

MATERIALS AND METHODS

The experiment was conducted at the department of Physiology and Pharmacology, Hajee Mohammad Danesh Science & Technology University, Dinajpur, during the period from January to June/2014. To complete the research work following steps were followed:

3.1 Collection of the plant and spices:

Some plants are collected from the HSTU campus, Dinajpur and spices sample was collected from Bahadur Bazar Market, Dinajpur. Plants are Fern, Auram, Tobacco, spices are cardamon, Tezpat, Zira, Zowin, Masted oil, Small alach etc.

3.2 Collection and processing of plant material

Fern, Auram, Tobacco leaves, Cardamon, Tezpat, Zira, Zowin, Masted oil, Small alach were selected for effectiveness as thrombolytic agent on Cardiovascular system. Mature and disease free Fern, Auram, Tobacco leaves, Cardamon, Tezpat, Zira, Zowin, Masted oil, Small alach leaves were collected from HSTU campus.

3.3 Preparation of Ethanol Extract:

3.3.1 Ethanol

Ethanol also called ethyl alcohol pure alcohol, grain alcohol, or drinking alcohol, is a volatile, flammable, colorless liquid with the structural formula $\text{CH}_3\text{CH}_2\text{OH}$, often abbreviated as $\text{C}_2\text{H}_5\text{OH}$ or $\text{C}_2\text{H}_6\text{O}$. Ethanol is a psychoactive drug and is one of the oldest recreational drugs still used by humans. Ethanol can cause alcohol intoxication when consumed. Best known as the type of alcohol found in alcoholic beverages, it is also used in thermometers, as a solvent, and as a fuel. In common usage, it is often referred to simply as alcohol or spirits.

Ethanol is a 2-carbon alcohol with the empirical formula $\text{C}_2\text{H}_6\text{O}$. Its molecular formula is $\text{CH}_3\text{CH}_2\text{OH}$. An alternative notation is $\text{CH}_3\text{-CH}_2\text{-OH}$, which indicates that the carbon of a methyl group ($\text{CH}_3\text{-}$) is attached to the carbon of a methylene group ($\text{-CH}_2\text{-}$), which is attached to the oxygen of a hydroxyl group (-OH). It is a constitutional isomer of dimethyl ether.

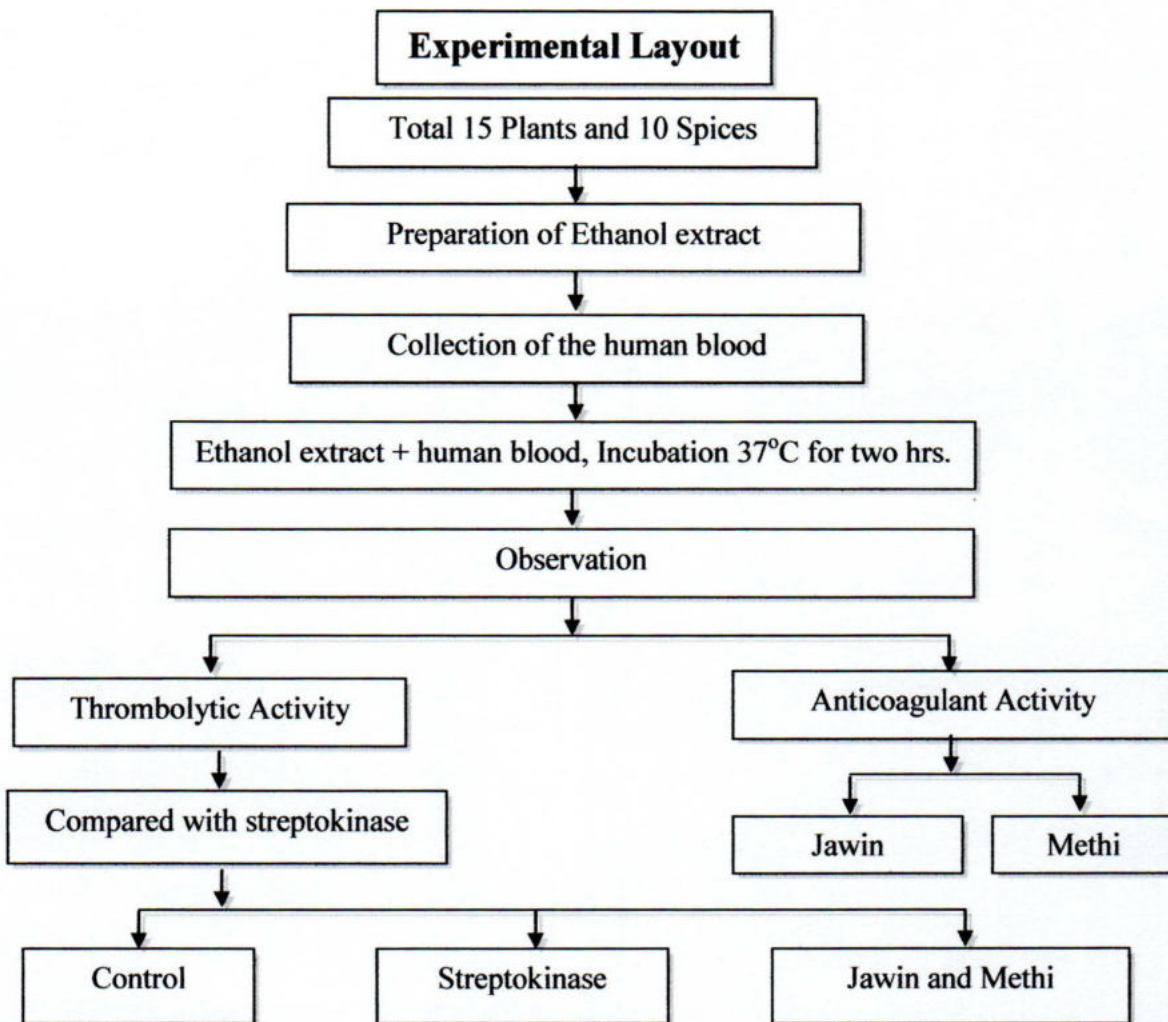


Fig. 3.1 Layout of the experiment

3.3.2 Extract

Extract is a solution of the essential constituents of a herbal agent. It is prepared by boiling the plant material in water and then evaporating the strained decoction to a desired concentration. Extracts harbour the more active principles of the medicinal plants, allowing the less active principles to be removed as a dross. Various solvents may be used including alcohol, water and glycerin. Extracts should always be prepared at the lowest temperature possible that is compatible with good extraction of the healing principles. If care is not taken, some of the lighter and more volatile oils and other important ingredients may be lost. In addition, the vessel in which the extract is prepared should always be tightly covered until such time as the extract has reached a sufficiently cool temperature (i.e., body temperature). Fluid Extract is an alcohol or glycerin preparation of herbal extract containing the active constituents in a definite ratio of plant material to solvent. There are several types of fluid extracts:

- Heated extracts are prepared by boiling (but preferably simmering) a herbal agent in water and then evaporating the strained decoction to a desired concentration — this yields a more concentrated herbal remedy.
- Fluid extract is made by evaporating an already prepared fluid (such as an infusion or decoction) to the desired concentration.
- Cold extract is similar to an infusion. It is prepared by taking twice as much herb as is desired for an infusion and then letting it sit in an enamel or non-metallic pot for eight to twelve hours. It is then strained and taken as one would take an infusion.
- Freeze drying is also used to get active ingredients out of plants and preserve them. The technique is used in the coffee industry as well as for herbs.
- Fresh Plant Extracts are becoming commercially popular. In these cases, fresh plants are used to make the tincture rather than dried material. Sometimes this results in a more potent preparation, other times less potent. The properties of a fresh plant extract will also vary from dried plant extracts, both positively and negatively.

3.3.3 Plant extract

A plant extract must, by definition, be obtained from a solid-liquid extraction. Solid-liquid extraction is defined as an operation to separate elements contained in a solid body

by solubilization with a solvent, and it may be followed by purification. The extract is contained in the solvent. If the solvent is an edible solvent, it is not necessary to dissociate it from the extract. If the solvent is not an edible solvent, separation allows obtaining a dry extract. Selection of raw materials, choice of solvent, process utilization and equipment performance are determining factors. These multiple parameters must be professionally combined. Only their proper combination allows obtaining high-quality plant extracts.

3.3.4 Preparation of leaves extract

For the preparation of extract, the leaves were dried in sun for 10 days and followed by oven at 55-60°C for 2 days. The dried leaves were pulverized with a blender. A 25 (unit) mesh diameter sieve was used to obtain the fine dust, after then dust was preserved in airtight plastic container until they were directly used for screening and preparation of water extract. 10g each leaves powder was added to 70ml of distilled water and was shaking overnight at room temperature, filtered and distilled water was added up to 100ml to make 10% extract.



Figure 1. Preparation of ethanol plant extract

3.3.5 Preservation of Plant Extract

The collected plant material (plant parts: stems, leaves, flowers, roots, bark etc.) is dried in a ventilated oven at 45 °C for 24 H, and subsequently milled to a fine powder by means of an IKA Universal Mixer M20 (or other type). An amount of 20.0 g of the dried plant powder is weighed in an Erlenmeyer of 100 ml to which 70 ml of hexane (purity grade 99 %) is added (the plant sample has to be submerged with solvent) for pre-

extraction. The Erlenmeyer is placed in a sonicator-bath (Branson 8210 or some other type) and sonicated at a temperature 40 °C during 30 minutes. The mixture is filtered using paper filter, followed by washing the Erlenmeyer with 20 ml of hexane and then with 50 ml of hexane. The filtrate is poured in a round-bottomed flask and the solvent is concentrated in vacuo (at about 11 mm Hg) up to 5-10 ml by means of rotavapor, utilizing a water bath at 40°C. This residue is brought in a 30-ml vessel to let the solvent evaporate. The open vessel is left overnight in a well-ventilated hood in order to evaporate the last traces of the solvent in the hexane pre-extract. The solids, collected on the filter, are broken up and dried in the air overnight in the hood. The dried material is extracted in the same way with methanol-water (90:10). The dried material from the filters placed in an Erlenmeyer of 100 ml to which 70 ml of 90 % methanol is added. The mixture is sonicated as above at 40C during 30 minutes, after which it is filtered, followed by washing the Erlenmeyer with 20 ml of 90 % methanol. The filtrate is poured in a round-bottomed flask and the solvent is evaporated in vacuo completely. The dry 90% methanol extract is dissolved in as little as possible 100 % methanol by using the sonicator-bath and poured in a 30-ml vessel to let it evaporate overnight in the hood.

3.4 Collection and management of Human Blood

10ml Human blood were collected from Vein by 10ml syringe. This blood were kept into 20 micropipet. Each micropipet contain 0.5ml blood. After few minutes blood automatically was cloted and serum was separated from the tube. After then blood containing pipet kept into pipet stand.

3.5 Different plant extract mixing with clotted blood

100 micromililitre ethanol plant extract measured by micropipet syringe and mixed with specific plant extract with specific test tube. Three blood test were preserved for control.

3.6 Streptokinase (SK)

3.6.1 Streptokinase (SK) is an enzyme secreted by several species of streptococci that can bind and activate human plasminogen. SK is used as an effective and inexpensive thrombolysis medication in some cases of myocardial infarction (heart attack) and pulmonary embolism. Streptokinase belongs to a group of medications known as fibrinolytics, and complexes of streptokinase with human plasminogen can hydrolytically activate other unbound plasminogen by activating through bond

cleavage to produce plasmin. There are three domains to Streptokinase, denoted α (residues 1–150), β (residues 151–287), and γ (residues 288–414). Each domain binds plasminogen, although none can activate plasminogen independently.

3.6.2 Mechanism of action

Plasmin is produced in the blood to break down fibrin, the major constituent of blood thrombi, thereby dissolving clots once they have fulfilled their purpose of stopping bleeding. Extra production of plasmin caused by streptokinase breaks down unwanted blood clots, for example, in the lungs (pulmonary embolism). The usual activation of Plasminogen (Plgn) is by proteolysis of the Arg561—Val562 bond. The amino group of Val562 then forms a salt-bridge with Asp740, which triggers a conformational change producing the active protease Plasmin (Pm). When (SK) is present, it binds to Plgn to form a complex (SK. Plgn) that converts substrate Plgn to Pm. Residues 1–59 of SK regulate its capacity to induce an active site in bound Pg by a nonproteolytic mechanism and to activate substrate Pg in a fibrin-independent manner. This complex subsequently rearranges to an active complex although the Arg561–Val562 bond remains intact. Therefore another residue must substitute for the free amino group of Val562 and provide a counterion for Asp740 in this active complex. Two candidates for this counterion have been suggested: Ile1 of streptokinase and Lys698 of Plgn. Deletion of Ile1 of SK markedly inhibits its capacity to induce an active site in plasminogen, which supports the hypothesis that establishment of a salt bridge between Ile1 of SK and Asp740 of plasminogen is necessary for SK to induce an active site in plasminogen by a nonproteolytic mechanism. In contrast with the Ile1 substitutions, the Lys698 mutations also decreased the dissociation constant of the SK complex by 15 to 50 fold. These observations suggest that Lys698 is involved in formation of the initial SK. Plgn complex.

3.7 Plant material collection

Carum copticum (Ajowan) and *Triconella foenum-graecum* (Methi/Fenugreek) was collected from local market Bahadur Bazar at Dinajpur in Bangladesh. The seed were washed with fresh water and dried under shade at room temperature. The seed were sinked into ethanol at two days. The seed were grinded wit mortar and pestle and squeezed out the liquid portion with hand and centrifuged at 1000 rpm. Then dried out

the water from the centrifugal material and powdered and stored. 60g of powdered drug was extracted separately with methanol, petroleum ether by continuous hot percolation in soxlet apparatus and with water by cold maceration for 3 days respectively. All the extracts were filtered and evaporated using a rotary evaporator. Dried extracts were stored at 20°C until used.

3.8 Study population

Blood samples obtained from my dear batchmates, were used to assess the anticoagulant effects of *Triconella foenum-graecum* (*Methi/Fenugreek*). Participants were 25-26 years old. They had been chosen for this study according to the following criteria: having normal prothrombin time, not suffering from any cardiovascular diseases (hypertension, congestive heart failure, coagulation disorders such as, Hemophilia A or B) or diabetes, not recently using nonsteroidal anti inflammatory drugs, not obese or smokers and free from dyslipidemic disorders.

3.9 Collection of blood samples

The blood samples were obtained from normal individuals by using sterile syringes, withdrawn from vein of right arm of each individual and placed separately in containers containing tri-sodium citrate to prevent the clotting process. Centrifugation (15 minutes at rate 3000 rpm) was carried out to separate the blood cells from plasma in order to obtain pure platelet plasma (ppp) for prothrombin time test. The obtained plasma sample of each individual were poured separately in plane containers using automatic pipette and stored at room temperature.

3.10 Invitro thombolytic activity of *Carum copticum* (*Ajowan*)

Blood sample: Blood (n=6) was drawn from healthy human volunteers without a history of oral contraceptive or anticoagulant therapy and 1ml of blood was transferred to the previously weighed micro centrifuge tubes and was allowed to form clots.



Figure 2. Lysis of the clotted blood (Ajawin)

Thrombolytic activity: The thrombolytic activity of all extracts was evaluated by the method developed by Daginawala (2006) and slightly modified by Kawsar *et al.* (2011) using streptokinase (SK) as the standard.



Figure 3. Control



Figure 4. Blood Lysis by Streptokinase

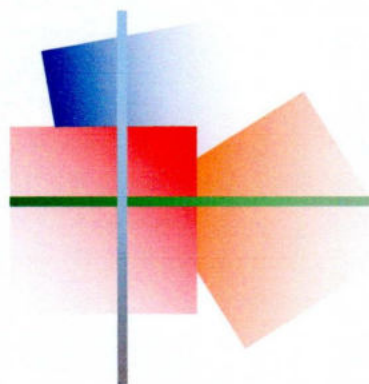
3.11 Invitro anticoagulant activity of *Triconella foenum-graecum* (Methi)

3.11.1 Phytochemical screening

Dried extracts were subjected for the presence of different phytoconstituent like alkaloid, steroid, flavonoid, tannin, glycoside etc.

3.12.2 Collection of blood and Plasma re-calcification

0.2 ml plasma, 0.1 ml of crude extract of different concentration and different volume of CaCl_2 (25 mM) were added together in a clean fusion tube and incubated at 37°C in water bath. For control experiment extract solution was replaced by same volume of 0.9% saline water. The clotting time was recorded with stopwatch by tilting the test tubes every 5 seconds. This time is called the prothrombin time.



Chapter 4

RESULTS AND DISCUSSION

CHAPTER 4

RESULTS AND DISCUSSION

This experiment was conducted to study the Thrombolytic activity of ethanol plant extract that may act on cardiovascular thrombus.

This experiment was held under the Department of Physiology & Pharmacology, Faculty of Veterinary and Animal Science.

Finally five plant and spices have thrombolytic activity after first trial. I was carrying on my experiment these five plant and spices such as ajawin, pesta, fern, auram (kochu) and Tobacco plant.

The experimental units were kept in the laboratory. At first I weighted micro test tubes. The weight of each test tube is 0.75g. Fresh and clean water was made available at all the times. The experiment was conducted according to the completely randomized design.

Thrombolytic activity: As a part of discovery of cardio-protective drugs from natural sources the extractives of *Carum copticum* (*Ajowan*) were assessed for thrombolytic activity and the results are presented in Table 1. Addition of 100 μ l SK, a positive control (30,000 I.U.), to the clots and subsequent incubation for 90 minutes at 37°C, showed 91.67% lysis of clot. At the same time, distilled water was treated as negative control which exhibited negligible lysis of clot (4.74%). In this study, the carbon tetrachloride soluble fraction (CCSF) exhibited highest thrombolytic activity (57.33%).

Table 1: Thrombolytic property of different group

Sl. No.	Weight of Blood Before Lysis (gm) (mean \pm SEmean)	Weight of blood (After lysis) (mean \pm SEmean)	Level of Significance (P value)
Aajawin	0.3967 b \pm 0.03811	0.2200 \pm 0.1581	P > 0.001 1% Level of Significance
Methi	0.3967 b \pm 0.03811	0.2320 \pm 0.01304	
Streptokinase	0.3967 b \pm 0.03811	0.0320 \pm 0.01095	

Note: Values followed by same superscripts in the same Column are not statistically significant ($P > 0.001$), different superscripts indicate that difference is significant ($p < 0.05$). In this other tables, A=Total amount of blood, B= Weight of blood (After lysis), C= Result (amount of lysed blood) gm, Control no change.

Table 1 Revealed that

Group A initial weight 0.3967 final weight 0.2200 weight lose 0.1767.

Group B initial weight 0.3967 final weight 0.2320 weight lose 0.1647.

Group C initial weight 0.3967 final weight 0.0320 weight lose 0.3647.

The thrombolytic activity presents significantly ($p > 0.001$) in *Carum copticum* (Ajowan) than *Triconella foenum-graecum* (Methi).

Table 2: Lysis of clotted blood by Streptokinase at standard Thrombolytic agent

Sl. No.	Weight of Blood Before Lysis (gm)	Weight of blood (After lysis)	Result (amount of lysed blood) gm	Percentage	Average
1 st Day	0.39	0.02	0.378	94%	91.67%
2 nd Day	0.44	0.04	0.40	90%	
3 rd Day	0.45	0.04	0.41	91%	

Table 3: Comparison of Thrombolytic Property between aajawin and methi regarding streptokinase.

Treatment Group	N	Subset for alpha = 0.05	
		1	2
Streptokinase	5	.0320	
Aajawin	5		.2200
Methi	5		.2320
Sig.	1% level of significance		

Note: Values followed by same superscripts in the same Column are not statistically significant ($p > 0.001$), different superscripts indicate that difference is significant ($p < 0.001$). In this other tables, A=Total amount of blood, B= Weight of blood (After lysis), C= Result (amount of lysised blood) gm, Control no change.

Table 3 Revealed that

Group A initial weight 0.3680 final weight 0.1480 weight lose 0.2200.

Group B initial weight 0.3860 final weight 0.1540 weight lose 0.2320.

Group C initial weight 0.3820 final weight 0.1600 weight lose 0.2220.

The thrombolytic activity presents significantly ($p > 0.05$) in *Triconella foenum-graecum* (Methi/Fenugreek. less than *Carum copticum* (Ajowan)

Table 4: Thrombolytic activity of different fractions of *Carum copticum*

Sample	Thrombolytic Activity (% of lysis)
SK	91.67%
Water	4.74%
ECSF	57.33%

SK= Streptokinase, and CCSF= Ethanolic extract of carum capticum fraction of the seed extracts of *Carum copticum* (Ajowan)

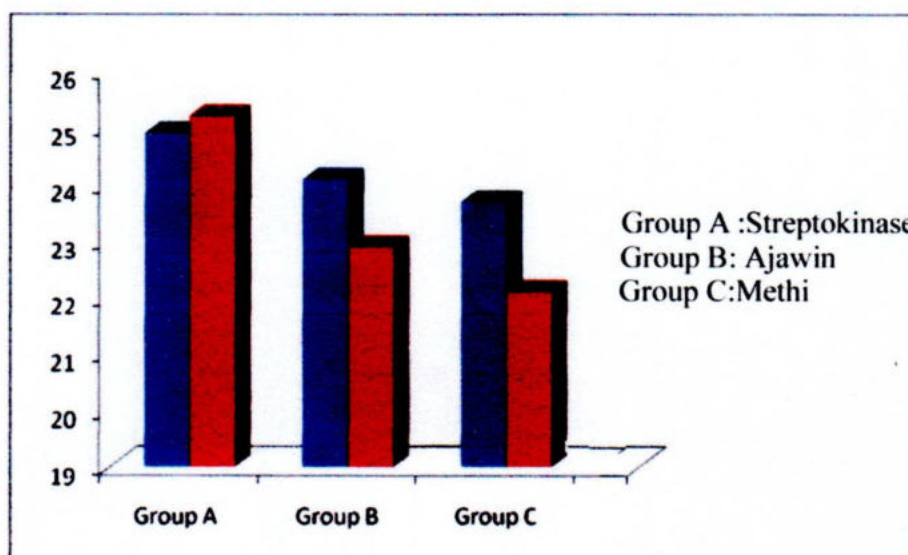


Fig. Thrombolytic activity

Table 5: Anticoagulant Property in Methi

Sl. No.	Clotting time (Minutes)	Control
A=1 st Day	56.40 a ± 0.8124	68.00 a ± 2.2583
B=2 nd Day	56.80 a ± 0.8602	70.00 a ± 0.4472
C=3 rd Day	57.80 a ± 0.8602	70.00 a ± 0.3162
Probability value	0.00	0.00
LSD value	2.603	4.134
CV %	3.31	4.33

Note: Values followed by same superscripts in the same Column are not statistically significant ($p > 0.05$), different superscripts indicate that difference is significant ($p < 0.05$). In this other tables, A=Total amount of blood, B= Weight of blood (After lysis), C= Result (amount of lysised blood) gm, Control no change.

Table 5 revealed that

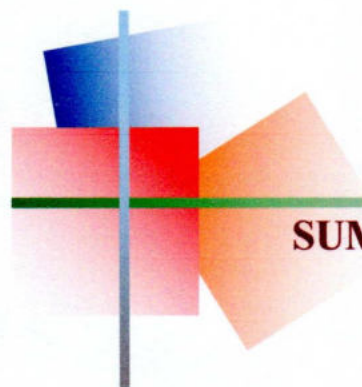
Group A Clotting time (Minutes) 56.40 control 68.00 .

Group B Clotting time (Minutes) 56.80 control 70.00 .

Group C Clotting time (Minutes) 57.80 control 70.00.

The anticoagulant activity presents significantly ($p > 0.05$) in *Triconella foenum-graecum* (Methi/Fenugreek).

Anti-coagulant activities of aqueous and methanolic extract of *Triconella foenum-graecum* (Methi/Fenugreek) were carried out. From the present study it is proved that both the extract have remarkable anti-coagulant activity than the control solution. The anticoagulant of methi is 1 hour 3 minutes. Further study is under progress to isolate the pure component fraction.



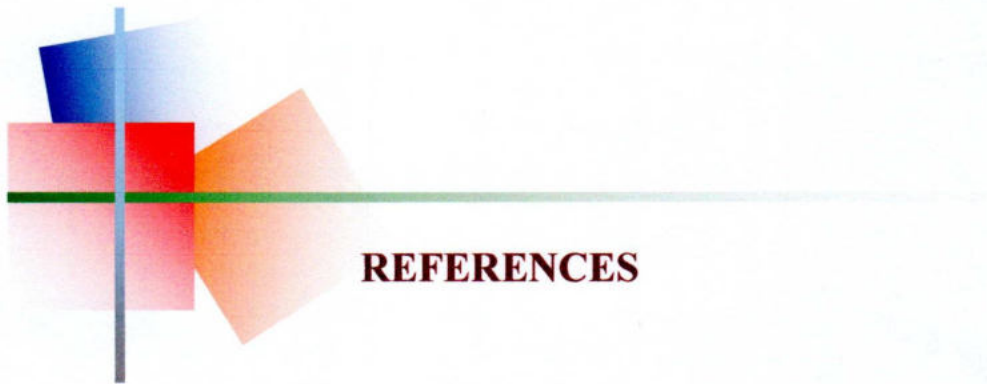
Chapter 5

SUMMARY AND CONCLUSION

CHAPTER 5

SUMMARY AND CONCLUSION

Five tube contain blood in three days experiment carried out in the laboratory in this research work. In this experiment extract of methi and ajawin were studied in terms of thrombolytic and anti-coagulant activity because we showed that this harbal plant or spices are available cost effective and produced treatment free from synthetic drug resudal effect suitable for human. In this research work extract of ajawin produced significant ($p < 0.05$) increased of treatment of cardiovascular system in the live body. This experiment also proved that anti-coagulant property of methi increased significantly ($p < 0.05$) that would be treated to cardiovascular system. No significant ($p > 0.05$) the anti-coagulant property of ajawin. The further studies are necessary to see any adverse effect in relation to histopathology before making a definite conclusion.



REFERENCES

REFERENCES

- A.B.P. Montanher, M.G. Pizzolatti, I.M.C. Brighente (2002).** "An Application of the Brine Shrimp Bioassay for General Screening of Brazilian Medicinal Plants", in *Acta Farm. Bonaerense*, vol. 21 no. 3. pp.175-178
- A.K. Anwar, M. Ashfaq, and M.A. Nasveen (1979).** *Pharmacognostic Studies of Selected Indigenous Plants of Pakistan*, Pakistan Forest Institute, Peshawar NWFP, Pakistan, pp. 15-35.
- A.K. Duttaroy, (2007).** "Taking tomatoes to heart", in *Advances in Medicinal Plant Research*. vol. 37 no. 661 India: Research Signpost. pp. 1-10
- A.S. Gallus (1998).** Thrombolytic therapy for venous thrombosis & pulmonary embolism. *Bailliere's Clin. Haematol.* 11: 663-673.
- A.V. Krishnaraju, T.V.N. Rao, D. Sundararaju, M. Vanisree, H.S. Tsay, and G.V. Subbaraju (2006).** Biological screening of medicinal plants collected from Eastern Ghats of India using *Artemia salina* (brine shrimp test), *Int. J. Appl. Sci. Eng.* 4: 115-125.
- Adams D.S., Griffin L.A, Nachajko W.R., Reddy V.B., Wei C.M. (1999).** A synthetic DNA encoding a modified human urokinase resistant to inhibition by serum plasminogen activator inhibitor. *J Biol Chem*; 266: 8476-8482.
- Aderem, A. and Ulevitch, R. J. (2000).** Toll-like receptors in the induction of the innate immune response. *Nature.* 406, 782-787.
- Aubréville, A. (1960).** Flore du Cambodge du Laos et du Viet Nam. Myanmar Medicinal Plant Database Harbalpedia. 2010. issue. 407, 785-789.
- Ayafor, J. F. (1972).** Limonoids and phytol derivatives from *Cedrela sinensis*. *Tetrahedron.* 28: 9343.

- Ayafor, J.F. (1972).** Limonoids and phytol derivatives from *Cedrela sinensis*. *Tetrahedron*. **28**, 9343.
- B.N. Tochi, Z. Wang, S. Xu, W. Zhang (2008).** "Therapeutic Application of Pineapple Protease (Bromelain): A Review", in *Pakistan Journal of Nutrition*. vol. 7 no. 4. Asian Network for Scientific Information. pp. 513-520
- Bauer, A.W.; Kirby, W. M. M., Sherris, J. C. and Tuck, M. (1966).** Antibiotic susceptibility testing by a standardized disc diffusion method. *Am. J. Clin. Pathol.* **45**: 493-496.
- Bazzano LA, He J, Ogden LG, Loria CM, Vupputuri S, Myers L, Whelton PK (2002).** Fruit and vegetable intake and risk of cardiovascular disease in US adults: the first National Health and Nutrition Examination Survey Epidemiologic Follow-up Study. *Am J Clin Nutr* , **76**:93-99.
- Bell, E., Creelman, R.A., and Mullet, J.E. (1995).** A chloroplast lipoxygenase is required for wound-induced jasmonic acid accumulation in *Arabidopsis*. *Proc. Natl. Acad. Sci. USA* **92**, 8675-8679.
- Bessire, M., Chassot, C., Jacquat, A.C., Humphry, M. and Borel, S., Petetot, J.M.C., Metraux, J.P. and Nawrath, C. (2007).** A permeable cuticle in *Arabidopsis* leads to a strong resistance to *Botrytis cinerea*. *EMBO J.* **26**, 2158-2168.
- Bhide, MB. (1977).** Studies on the antiasthmatic activity of *Mesua ferrea*. *Haff. Inst.* **5**: 27.
- Briggs WH, Folts JD, Osman H.E. (2001).** Administration of onion inhibits platelet-mediated thrombosis in dogs. *J Nutr*; **131**(10): 2619-2622.
- Briggs WH, Folts JD, Osman HE (2001).** Administration of raw onion inhibits platelet-mediated thrombosis in dogs. *J Nutr*, **131**(10):2619-2622.
- Cameron, D. G. (1988).** Tropical and subtropical pasture legumes. *Queensland Agricultural Journal*. March-April:110-113.

- Chakma J.S., Rahman M.A., Islam S., Rana M.S., Ahmed N.U. (2011).** Analgesic and anti-inflammatory effect of *Clausena suffruticosa* root extract in animal model. *J Sci Res*, **3**:631-639.
- Chakravarty, A.K., Ghosh, P., Pratim, D. and Pakrashi, SC. (1982).** Simple aromatic amines from *Justica gendarussa*. 13 C-NMR spectra of the bases and their analogues. *J Sci Res*, **4**:625-635.
- Chassot, C., Nawrath, C. and Metraux, J. P. 2007.** Cuticular defects lead to full immunity to a major plant pathogen. *Plant J*. **49**, 972-980.
- Chew AL, Jessica JJA, Sasidharan S (2012).** Antioxidant and antibacterial activity of different parts of *Leucas aspera*. *Asian Pac J Trop Biomed*, **2**(3):176-180.
- Chinchilla, D., Zipfel, C., Robatzek, S., Kemmerling, B., Nürnberger, T., Jones, J.D., Felix, G. and Boller, T. (2007).** A flagellin-induced complex of the receptor FLS2 and BAK1 initiates plant defence. *Nature*. **26**, 497-500.
- Collen D (1990).** Coronary thrombosis: streptokinase or recombinant tissue-type plasminogen activator? *Ann Intern Med*; **112**: 529-538.
- D. Collen (1996).** Fibrin-selective thrombolytic therapy for acute myocardial infarction, *Circulation* **93**: 857-865.
- D. Prashar, S.L. Khokra, R. Purohit, S. Sharma (2011).** "Curcumin: A Potential Bioactive Agent", in *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. vol. 2 no. 4. pp. 46
- D.B. Baruah, R.N. Dash, M.R. Chaudhari, and S.S. Kadam (2006).** Plasminogen activators: A comparison, *Vascular Pharmacol*. **44**: 1-9.
- D.S. Rani and K. Nand. (2004).** "Ensilage of pineapple processing waste for methane generation", in *Waste Management*. vol. 24 no. 5. New York. pp. 523-528
- Daginawala, H. F.; Prasad, S., Kashyap, R. S., Deopujari, J. Y., Purohit, H. J. and Taori, G. M. (2006).** Development of an *in vitro* model to study clot lysis activity of thrombolytic drugs. *Thrombosis J*. **4**: 14.

- Daginawala, HF., Prasad, S., Kashyap, RS., Deopujari, JY., Purohit, HJ. and Taori, GM. (2006).** Development of an in vitro model to study clot lysis activity of thrombolytic drugs. *Thrombosis Journal.* 4: 14.
- De Smet, I., Vassileva, V., De Rybel, B., Levesque, M. P., Grunewald, W., Van Damme, D., Van Noorden, G., Naudts, M., Van Isterdael, G., De Clercq, R., Wang, J.Y., Meuli, N., Vanneste, S., Friml, J., Hilson, P., Jürgens, G., Ingram, G.C., Inzé, D., Benfey, P. N. and Beeckman, T. (2008).** Receptor-like kinase ACR4 restricts formative cell divisions in the *Arabidopsis* root. *Science.* 322, 594-597.
- Deka, R. K. and Sarkar, C. R. (1990).** Nutrient composition and anti-nutritional factors of *Dolichos lablab* L seeds. *Food Chemistry.* 38:239-246.
- Demrow H.S., Slane P.R., Folts J.D. (1995).** Administration of wine and grape juice inhibits in vivo platelet activity and thrombosis in stenosed canine coronary arteries. *Circulation;* 91(4): 1182-1188.
- Demrow H.S., Slane P.R., Folts J.D. (1995).** Administration of wine and grape juice inhibits *in vivo* platelet activity and thrombosis in stenosed canine coronary arteries. *Circulation,* 91(4):1182-1188.
- E.M. El-Sayed, Amal S.A. El-azeem, A.A. Afify, M.H. Shabana, and H.H. (2011).** Ahmed. Cardioprotective effects of *Curcuma longa* L. extracts against doxorubicin-induced cardiotoxicity in rats, *J. Med. Plants Res.* 5: 4049-4058.
- F. Ferreres, M. Taveira, D.M. Pereira, P. Valentao and P.B. Andrade (2010).** "Tomato (*Lycopersicon esculentum*) Seeds: New Flavonols and Cytotoxic Effect", in *Journal of Agricultural and Food Chemistry.* vol 58 no. 5. American Chemical Society. pp. 2854-2861.
- F.El Sabban, (2009).** "Garlic as an antithrombotic and antiplatelet aggregation agent", in *Journal of Chinese Clinical Medicine.* vol.4 no. 5. Kuwait. pp. 288-292.

- Finnish National Public Health Institute (2002).** "Recommendation for indicators, international collaboration, protocol and manual of operations for chronic disease risk factor surveys". Finland. **06(3):364-7.**
- Fowles RG, Mootoo BS, Ramsewak RS, Khan A. (2012).** Toxicity-structure activity evaluation of limonoids from *Swietenia* species on *Artemia salina*. *Pharm Biol*, **50(2):264-7.**
- G. Persoone (1980).** Proceeding of International Symposium on Brine Shrimp, *Artemia salina*, Vol. 1-3. Universal Press. Wilteren, Belgium. 586: 250-260.
- G.C. Leta, P.A.S. Mourao, and A.M.F. Tovar (2002).** Human venous and arterial glycosaminoglycans have similar affinity for plasma low-density lipoproteins, *Biochim. Biophys. Acta.* 586: 243-253.
- Gajraj Sharma, E. Jayachandran, G.M. Sreenivasa (2008).** Synthesis of Bioactive Molecule FluoroBenzothiazole Comprising Quinazolylinyl Oxadiazole Derivative For Biological and Pharmacological Screening, *Ultra Chemistry* vol.4(1); 103: 114.
- Gelias & M.N.A. Rao (1998).** Inhibition of Albumin Denaturation and Antiinflammatory Activity of Dehydrozingerone and Its Analogs, *Indian. J. Exp. Biol*, vol. 26, July.
- Gesler WM. (1992).** Therapeutic landscapes: medical issues in light of the new cultural geography. *Soc Sci Med*; 34: 735-746.
- Gifford, M.L., Dean, S. and Ingram, G.C. (2003).** The *Arabidopsis ACR4* gene plays a role in cell layer organisation during ovule integument and sepal margin development. *Development* **130**, 4249-4258.
- H. R. Maurer (2001).** "Bromelain: Biochemistry, pharmacology and medicinal use", in *Cellular and Molecular Life Sciences*. vol. 58. pp. 1231-1245.

- H.S. Demrow, P.R. Slane, and J.D. (1995).** Folts. Administration of wine and grape juice inhibits *in vivo* platelet activity and thrombosis in stenosed canine coronary arteries, *Circulation* 91:1182-1188.
- Haque, M. A.; Islam, A. S. M. and Shahriar, M. (2011).** Antimicrobial, Cytotoxicity and Antioxidant Activity of *Tinospora crispa*. *Journal of Pharmaceutical and Biomedical Sciences*. 12(13): 1-4.
- Harborne, J. B. (1998).** Phytochemical methods. A guide to modern techniques of plant analysis. 3rd edn., Chapman and Hall int Ed., New York.
- Hossain, M. I.; Sharmin, F. A., Akhter, S., Bhuiyan, M. A. and Shahriar, M. (2012).** Investigation of cytotoxicity and *In-vitro* antioxidant activity of *Asparagus racemosus* root extract. *International Current Pharmaceutical Journal*. 1(9):250-257.
- I. Chattopadhyay, K. Biswas, U. Bandyopadhyay and R.K. Banerjee (2003).** "Turmeric and curcumin: Biological Actions and Medicinal Applications", in *Current Science*. vol.87 no. 1. pp. 44-48.
- I. Makoto, K., et al. (2003).** "Identification of Six Phenylpropanoids from Garlic Skin as Major Antioxidants", *Journal of Agricultural and Food Chemistry*. vol. 51 no. 25. New Jersey. pp. 7313-7317.
- I. Mohantya, D.S. Aryaa, A. Dindab, S. Joshia, K.K. Talwarc, and S.K. Guptaa. (2004).** Protective effects of *Curcuma longa* on ischemia-reperfusion induced myocardial injuries and their mechanisms, *Life Sci*. 75: 1701-1711.
- I.N. Khan Md., R. Habib Md., M. Rahman, A. Mannan Md., M.I. Sarker and S. Hawlader (2011).** "Thrombolytic potential of *Ocimum sanctum* L., *Curcuma longa* L., *Azadirachita indica* L. and *Anacardim occidentale* L.", in *Journal of Basic and Clinical Pharmacy*, vol.2 no. 3. Bangladesh. pp. 125-127.
- Islam MS, Rahman M.A. (2013).** Phytochemicals of the whole *Leucas aspera* extract show potent antioxidant, antibacterial and cytotoxic effects. *Asian Pac J Trop Biomed*, 3:273-279.

- Ita SO, Etim OE, Ben EE, Ekpo O.F. (2007).** Haematopoietic properties of ethanolic extract of *Ageratum conyzoides* (goat weed) in albino rats. *Niger J Physiol Sci*, 22(1-2):83-7.
- J. Yamamoto, et al. (2003).** "Tomatoes have natural anti-thrombotic effects", in *British Journal of nutrition*. vol. 90. pp. 1031-1038.
- J. Yamamoto, K. Yamada, A. Naemura, T. Yamashita, and R. Arai (2005).** Testing various herbs for antithrombotic effect, *Nutrition* 21: 580-587.
- J.A. Hoekstra (1987).** "Acute bioassays with control mortality", in *Water, Air & Soil Pollution*. vol. 35 no. 3-4. D. Reidel Publishing Company. pp. 311-317.
- J.M. Wardlaw, E. Berge, G. del Zoppo, and T. Yamaguchi T. (2004).** Thrombolysis for acute ischemic stroke, *Stroke* 35: 2914-2915.
- Jang I.K., Gold H.K., Ziskind A.A., Fallon J.T., Holt R.E., Leinbach R.C., May J.W., Collen D. (1989).** Differential sensitivity of erythrocyte-rich and platelet-rich arterial thrombi to lysis with recombinant tissue-type plasminogen activator. *Circulation*, 79:920-928.
- Jennings K:** Antibodies to streptokinase –once is enough. *BMJ* 1996; 312: 393-394.
- Jennings K:** Antibodies to streptokinase – once is enough. *BMJ* 1996, 312:393-394.
- K.J. Joshipura, A. Ascherio, J.E. Manson, M.J. Stampher, E.B. Rimm, and F.E. Speizer (1999).** Fruit and vegetable intake in relation to risk of ischemic stroke, *JAMA* 282: 1233-1239.
- Kawsar, M. H.; Sikder, M. A., Rana, M. S., Nimmi, I. and Rashid, M. A. (2011).** Studies of Thrombolytic and cytotoxic properties of two asteraceous plants of Bangladesh. *Bang. Pharm. J.* 14: 103-106.
- Kay, D. E. (1979).** **Hyacinth Bean** - Food Legumes. *Crop and Product Digest* No. 3. Tropical Products Institute. 16:184-196.

- Koffuor G.A., Amoateng P., Andey T.A. (2011).** Immunomodulatory and erythropoietic effects of aqueous extract of the fruits of *Solanum torvum* Swartz (Solanaceae). *Pharmacog Res*, 3(2):130-4.
- Kokate, C. K. (2001).** Pharmacognosy 16th edn. Nirali prakashani, Mumbai India.
- Konwarh, R., Kalita, D., Mahanta, C., Mandal, M. and Karak, N. (2010).** Magnetically recyclable, antimicrobial, and catalytically enhanced polymer-assisted "green" nanosystemimmobilized *Aspergillus niger* amyloglucosidase. *Appl. Microbiol. Biotechnol.* 87: 1983 - 1992.
- Kowalski M., Brown G., Bieniasz M, Oszejca K., Chabielska E., Pietras T., Szemraj Z., Makandjou-Ola E., Bartkowiak J., Szemraj J. (2009).** Cloning and expression of a new recombinant thrombolytic and antithrombotic agent - a staphylokinase variant. *Acta Biochem Pol*, 56:41-53.
- L. Zhiguang, W. Hongli, L. Jiazeng, G. Zhang, and C. Gao (2000).** Basic and clinical study on the antithrombotic mechanism of glycoaminoglycan extracted from sea cucumber, *Chin. Med. J.* 113: 706-711.
- L.A. Bazzano, J. He, L.G. Ogden, C.M. Loria, S. Vupputuri, L. Myers, and P.K. Whelton (2002).** Fruit and vegetable intake and risk of cardiovascular disease in US adults: the first national health and nutrition examination survey epidemiologic follow-up study, *Am. J. Clin. Nutr.* 76: 93-99.
- LAD Williams, A Connar, L Latore et al. (2008).** The invitro Antidenaturation Effects Induced by Natural Products and Nonsteroidal Compounds in Heat Treated (Immunogenic)Bovine Serum Albumin is Proposed as a Screening Assay for the Detection of Anti-inflammatory Compounds' , without the use of Animals ,in the early Stages of the Drug Discovery Process, *West Indian Med J.*; 57(4): 327.
- Lee HS:**How Safe is the readministration of streptokinase drug safe. 1995; 13: 76-80.
- Leta G.C., Mourão P.A., Tovar A.M. (2002).** Human venous and arterial glycosaminoglycans have similar affinity for plasma low density lipoprotein. *Biochim Biophysics Acta*; 1586(3): 243- 245.

- Leta G.C., Mourão P.A., Tovar A.M. (2002).** Human venous and arterial glycosaminoglycans have similar affinity for plasma low-density lipoproteins. *Biochim Biophys Acta*, 1586(3):243-253.
- Licciardi P.V., Underwood J.R. (2011).** Plant-derived medicines: a novel class of immunological adjuvants. *Int Immunopharmacol*, 11(3):390-8.
- Lijnen H.R., Vanhoef B., DeCock F., Okada K., Ueshima S., Matsuo O. (1991).** On the mechanism of fibrin – specific plasminogen activation by staphylokinase. *J Biol Chem*; 266: 11826-11832.
- Liu S., Manson J.E., Lee I.M., Cole S.R., Hennekens C.H., Willett W.C., Buring J.E. (2000).** Fruit and vegetable intake and risk of cardiovascular disease: the Women's Health Study. *Am J Clin Nutr* , 72:922-928.
- Logarto P.A., Silva Y.R., Guerra S.I., Iglesias B.L. (1997).** Comparative study of the assay of *Artemia salina* L. and the estimate of the medium lethal dose (LD₅₀ value) in mice, to determine oral acute toxicity of plant extracts. pp. 256, 265.
- M. C. Cantoria (1994).** Selected Topics in Pharmacognosy, Taguig: National Academy of Science and Technology, pp. 417, 430
- M. Chethankumar, N. Anand, N.S. Gangadhara (2010).** “Isolation and characterization of an antioxidant protein from turmeric (*Curcuma longa* L.) peel waste: A new biological source”, in Journal of Pharmacy Research. LA: Association of Pharmaceutical Innovators. Issue 11. pp. 2659-2662.
- M.A. DeWood, J. Spores, R. Notske, L.T. Mouser, R. Burroughs, M.S. Golden, and H.T. Lang (1980).** Prevalence of total coronary occlusion during the early hours of transmural myocardial infarction, *N. Engl. J. Med.* 303: 897-902.
- M.J. Davies, and A.C. (1985).** Thomas. Plaque fissuring: the cause of acute myocardial infarction, sudden ischaemic death, and crescendo angina, *Br. Heart J.* 53: 363-373.

- M.S. Lenucci, et al. (2009).** "Carotenoid Content during Tomato (*Solanum Lycopersicum* L.) Fruit Ripening in Traditional and High-pigment Cultivars" in Italian Journal of Food Science. vol. 21 no. pp. 461-474.
- M.W. Gillman, L.A. Cupples, D. Gagnon, B.M. Posner, R.C. Ellison, W.P. Castelli, and P.A. (1995).** Wolf. Protective effect of fruits and vegetables on development of stroke in men, *JAMA* 273: 1113-1117.
- Marder V.J. (1993).** Recombinant streptokinase opportunity for an improved agent. *Blood Coagulation Fibrinolysis*; 4: 1039-1040.
- Maundu, P. M.; Ngugi, G. W. and Kabuye, C. H. S. (Eds.) (1999).** "*Lablab purpureus*," In: Traditional Food Plants of Kenya, English Press Ltd., Nairobi. pp. 154-155.
- Meyer B.N., Ferrigni N.R., Putnam J.E., Jacobsen L.B., Nichols D.E., McLaughlin J.L. (1982).** Brine Shrimp: A convenient general bioassay for active plant constituents. *Planta Med*, 45:31-34.
- Milford, R. and Minson, D. J. (1968).** The effect of age and method of haymaking on the digestibility and voluntary intake of the forage legumes *Dolichos lablab* and *Vigna sinensis*. *Australian Journal of Experimental Animal Husbandry*. 8:409-418.
- Min S.K., Han S.M., Kim H.T., Kwon O.C., Lee S., Kim J.K. (2004).** Algal fucoidan, unlike heparin, has thrombolytic activity in a murine arterial thrombosis model. *Blood Coagulation Fibrinolysis*. 50:45-56.
- Mrunthunjaya, K. and Hukkeri, V.I. (2007).** Antioxidant and free radical scavenging potential of *J. gendarussa* Burm, leaves *in vitro*. *Natural Prod. Sci*. 3: 199-206.
- Mucklow J.C. (1995).** Thrombolytic treatment streptokinase is more economical than Alteplase. *BMJ*; 311: 1506.

- N. O’Kennedy, L. Crosbie, J. van Lieshout and A. Duttaroy (2006).** “Effects of antiplatelet components of tomato extract on platelet function in vitro and ex vivo: a time-course cannulation study in healthy humans” in *The American Journal of Clinical Nutrition*. vol. 84 no. 3. USA: American Society for Nutrition. pp.570-578.
- N. Rajapakse, W.K. Jung, E. Mendis, S.H. Moon, and S.K. Kim (2005).** A novel anticoagulant purified from fish protein hydrolysate inhibits factor XIIa and platelet aggregation, *Life Sci*. 76: 2607-2619.
- N.L. Ah Kee, N. Mnonopi, H. Davids, R.J. Naude, C.L. Frost, (2008).** “Antithrombotic/anticoagulant and anticancer activities of selected medicinal plants from South Africa”, in *African Journal of Biotechnology*, vol.7 no.3.
- N.L. Rozzi, R.K. Singh, R.A. Vierling and B.A. Watkins (2002).** “Supercritical Fluid Extraction of Lycopene from Tomato Processing Byproducts”, in *Journal of Agricultural and Food Chemistry*. vol.50 no. 9. Indiana: American Chemical Society. 2638-2643.
- N.P. Peteros and M.M. Uy (2010).** “Antioxidant and Cytotoxic Activities and Phytochemical Screening of Four Philippine Medicinal Plants”, in *Journal of Medicinal Plants Research*, vol.4 no. 5. Iligan City, Philippines: Mindanao State University. pp. 407-414
- Naderi G.A., Asgary S., Jafarian A., Askari N., Behagh A., Aghdam R.H. (2005).** Fibrinolytic effects of ginkgo biloba extract. *Exp Clin Cardiol*, 10(2):85-7.
- Nicolini F.A., Nichols W.W., Mehta J.L., Saldeen T.G., Schofield R., Ross M., Player D.W., Pohl G.B., Mattsson C. (1992).** Sustained reflow in dogs with coronary thrombosis with K2P, a novel mutant of tissue plasminogen activator. *J AM Coll Cardiol.*; 20: 228-235.
- Nurmburg, P.L., Knox, K.A., Yun, B.W., Morris, P.C., Shafiei, R., Hudson, A., and Loake, G.J. (2007).** The developmental selector AS1 is an evolutionarily conserved regulator of the plant immune response. *Proc. Natl. Acad. Sci. USA* 104, 18795-18800.

- O.L. Erukainure, J.A. Adejobi, R.O. Okafor, O.Y. Adenekan (2011).** “Protective effect of pineapple (*Ananas cosmosus*) peel extract on alcohol-induced oxidative stress in brain tissues of male albino rats”, in *Asian Pacific Journal of Tropical Medicine*. pp. 182-184.
- Okwuosa VN, Molta BS, Ebele S. (1993).** Toxicity of aqueous bark extracts of the tree *Balanitesa egyptiacaon* the fish *Oreochromis niloticus*. *Appl Parasitol*, 34(2):89-94.
- Orwa, C., Mutua, A., Kindt, R., Jamnadass, R. and Simons, A. 2009.** **Agroforestry Database. 4.0. P.F. Cavagnaro, A.C. Carmago, C.R. Galmarini and P.W. Simon (2007).** “Effect of Cooking on Garlic (*Allium sativum* L.) Antiplatelet Activity and Thiosulfonates Content”, in *Journal of Agricultural and Food Chemistry*. Wisconsin. pp.1280-1288.
- Pantzar M., Ljungh A., Wadström T. (1998).** Plasminogen binding and activation at the surface of *Helicobacter pylori* CCUG 17874. *Infect Immun*, 66(10):4976-4980.
- Parry M.A., Zhang X.C., Bode I. (2000).** Molecular mechanisms of plasminogen activation: bacterial cofactors provide clues. *Trends Biochem Sci*, 25(2):53-59.
- Pengelly, B. C. and Lisson, S. N. (2002).** “Strategies for Using Improved Forages to Enhance Production in Bali Cattle,” In: K. Entwistle and D. R. Lindsay, Eds., *Proceedings of a Workshop Australian Centre for International Agricultural Research (ACIAR)*, Bali, 4-7 February, pp. 29-33.
- Prasad S, Kashyap RS, Deopujari JY, Purohit HJ, Taori GM, Daginawala HF. (2006).** Development of an in vitro model to study clot lysis activity of thrombolytic drugs. *Thrombosis Journal*; 4: 14.
- Prasad S., Kashyap R.S., Deopujari J.Y., Purohit H.J., Taori G.M., Daginawala HF. (2007).** Effect of *Fagonia Arabica*(Dhamasa) on in vitro thrombolysis. *BMC Complem Altern M*, 7:36.

- Prasad S, Kashyap RS, Deopujari JY, Purohit HJ, Taori GM, Dagainawala HF (2006).** Development of an *in vitro* model to study clot lysis activity of thrombolytic drugs. *Thromb J*, 4:14.
- R. Srivastava, V. Puri, R.C. Srimal, and B.N. Dhawan (1986).** Effect of curcumin on platelet aggregation and vascular prostacyclin synthesis, *Arzneimittel forschung* 36: 715-717.
- Rahman M.A., Chakma J.S., Islam S., Ahmed N.U. (2011).** Evaluation of antioxidant, antibacterial, antifungal and cytotoxic effects of *Clausena suffruticosa* ethanolic extract. *J App Pharm Sci*, 1:90-95.
- Rahman M.A., Imran T. (2012).** Antioxidative, antimicrobial and cytotoxic effects of the phenolics of *Leea indica* (Burm. f.) Merr. leaf extract. *Saudi J Biol Sci.* 5: 56-70.
- Rahman MS., Sadhu SK., Hasan CM. (2007).** Preliminary antinociceptive, antioxidant and cytotoxic activities of *Leucas aspera* root. *Fitoterapia*, 78:552-555.
- Rahmatullah M., Mollik M.A.H., Ahmed M.N., Bhuiyan M.Z.A., Hossain M.M., Azam M.N.K., Seraj S., Chowdhury M.H., Jamal F., Ahsan S., Jahan R. (2010).** A survey of medicinal plants used by folk medicinal practitioners in two villages of Tangail district, Bangladesh. *American-Eurasian J Sust Agr*, 4(3):357-362.
- Raihan M.O., Habib M.R., Brishti A., Rahman M.M., Saleheen M.M., Manna M. (2011).** Sedative and anxiolytic effects of the methanolic extract of *Leea indica* (Burm. f.) Merr. leaf. *Drug Discov Ther*, 5(4):185-189.
- Rajapakse N., Jung W.K., Mendis E., Moon S.H., Kim S.K. (2005).** A novel anticoagulant purified from fish protein hydrolysate inhibits factor XIIa and platelet aggregation. *Life Sciences*; 76(22): 2607-2619.
- Rajapakse N., Jung W.K., Mendis E., Moon S.H., Kim S.K. (2005).** A novel anticoagulant purified from fish protein hydrolysate inhibits factor XIIa and platelet aggregation. *Life Sci*, 76(22):2607-2619.

- Rantnasooriya, W.D., Derianyagala, S.A. and Dehigas, D.C. (2007).** Antinociceptive activity and toxicological study of aqueous leaf extract of *J. gendarussa*. Phcog. Mag. 3: 145-155.
- Rios, J.L. and Recio, M.C. (2005).** Medicinal plants and Antimicrobial activity. *Journal of Ethnopharmacology*; 100: 80.
- Rouf S.A., Moo-Young M., Chisti Y. (1996).** Tissue plasminogen activator: characteristics, applications and production technology. *Biotechnol Adv*; 14: 239-266.
- S. Chanda, Y. Baravalla, M. Kaneria and K. Rakhollya, (2010).** "Fruit and Vegetable peels – strong natural source of antimicrobics", in Current Research, Technology and Education Topics in Applied Microbiology and Microbial Biotechnology. FORMATEX, pp. 445.
- S. Liu, J.E. Manson, I.M. Lee, S.R. Cole, C.H. Hennekens, W.C. Willett, and J.E. Buring (2000).** Fruit and vegetable intake and risk of cardiovascular disease: the women's health study, *Am. J. Clin. Nutr.* 72: 922-928.
- S. Prasad, R.S. Kashyap, J. Y. Deopujari, H. J. Purohit, G. M. Taori, and H. F. Daginawala (2007).** Effect of *Fagonia arabica* (Dhamasa) on *in vitro* thrombolysis, *BMC Complementary and Alternative Medicine* 7: 36. doi:10.1186/1472-6882-7-36.
- S. Prasad, R.S. Kashyap, J.Y. Deopujari, H.J. Purohit, G.M. Taori, and H.F. Daginawala (2006).** Development of an *in vitro* model to study clot lysis activity of thrombolytic drugs, *Thrombosis J.* 4: 14.
- S.A. Dahanukar, R.A. Kulkarni, N.N. Rege (2002).** "Pharmacology of Medicinal Plants and Natural Products", in Indian Journal of Pharmacology, vol. 32. 581-585.
- Shahriar, M. (2010).** Antimicrobial Activity of the Rhizomes of *Curcuma zedoaria*. *J. Bang. Academy of Sci.* 34 (2): 201-203.

- Shahriar, M. and Kabir, S. (2011).** Analgesic Activity of *Adiantum flabellulatum*. Dhaka Uni. J. Bio. Sci. 20 (1): 91-93.
- Shahriar, M.; Bahar, A.N.M., Hossain, M.I., Akhter, S., Haque M.A., and Bhuiyan, M.A. (2012).** Preliminary Phytochemical Screening, *In-Vitro* Antioxidant and Cytotoxic Activity of Five Different Extracts of *Withania somnifera* root. International Journal of Pharmacy. 2(3): 450-453.
- Shahriar, M.; Hossain, M. I., Bahar, A. N. M., Akhter, S., Haque, M. A., and Bhuiyan, M. A. (2012).** Preliminary Phytochemical Screening, *In-Vitro* Antioxidant and Cytotoxic Activity of Five Different Extracts of *Moringa Oleifera* Leaf. Journal of Applied Pharmaceutical Science. 2 (5): 65-68.
- Sikder, M.A., Kaiser, M.A., Parvez, M.M., Hossain, A.K.M. N., Akhter, F. and Rashid, M.A. (2011).** Preliminary Antimicrobial Activity and Cytotoxicity of Leaf Extracts of *Mesua nagassarium* (Burm.f.) Boletín Latinoam caribe Plant Med Aromat. 10 (1): 83 – 87
- Singh A., Singh S.K. (2005).** Molluscicidal evaluation of three common plants from India. *Fitoterapia*, 76(7-8):747-51.
- Skerman, P. J.; Cameron, D. G. and Riveros, F. (1991).** Leguminosas forrajeras tropicales. Colección FAO: Producción y Protección Vegetal, No. 2. Organización de las Naciones Unidas para la Agricultura y la Alimentación. Roma.
- Sofowora, A. (1993).** *Medicinal plants and Traditional Medicine in Africa*. Spectrum Books Limited, Ibadan.
- Srinivasan G.V., Sharanappa P., Leela N.K., Sadashiva C.T., Vijayan K.K. (2009).** Chemical composition and antimicrobial activity of the essential oil of *Leea indica* (Burm. f.) Merr. Flowers. *Nat Prod Rad*, 8:488-493.
- Stahl, Y., Wink, R.H., Ingram, G.C. and Simon, R. (2009).** A signaling module controlling the stem cell niche in Arabidopsis root meristems. *Curr. Biol.* 19, 909-914.

- Tang, D., Simonich, M. T. and Innes, R.W. (2007).** Mutations in LACS2, a long-chain acyl-coenzyme A synthetase, enhance susceptibility to avirulent *Pseudomonas syringae* but confer
- Tepe B., Daferera D., Sokmen M., Polissiou M., Sokmen A. (2004).** *In vitro* antimicrobial and antioxidant activities of the essential oils and various extracts of *Thymus eigi* M. Zohary et P.H. Davis. *J Agric Food Chem.*; 52: 1132-1137.
- Tetrahedron 38: 1797-1802. Marderosian, D., Giller, F.B. and Rola, F.C. (1976).** Phytochemical and toxicological screening of household ornamental plants potentially toxic to humans. *J. Toxicological. Environ. Health* 1: 939-953.
- Thomma, B.P.H.J., Eggermont, K., Penninckx, I.A.M.A., Mauch-Mani, B., Vogelsang, R., Cammue, B.P.A. and Broekaert, W.F. (1998).** Separate jasmonate-dependent and salicylate-dependent defence-response pathways in *Arabidopsis* are essential for resistance to distinct microbial pathogens. *Proc. Natl. Acad. Sci. USA* 95, 15107-15111.
- Tillet WS, Garner RL (1933).** The fibrinolytic activity of hemolytic streptococci. *J Exp Med*, 58:485-502.
- Torres-Urrutia C, Guzmán L, Schmeda-Hirschmann G, Moore-Carrasco R, Alarcón M, Astudillo L, Gutierrez M, Carrasco G, Yuri JA, Aranda E, Palomo I (2011).** Antiplatelet, anticoagulant, and fibrinolytic activity *in vitro* of extracts from selected fruits and vegetables. *Blood Coagul Fibrinolysis*, 22:197-205.

- Voisin, D., Nawrath, C., Kurdyukov, S., Franke, R.B., Reina-Pinto, J.J., Efremova, N., Will, I., Schreiber, L. and Yephremov, A. (2009).** Dissection of the complex phenotype in cuticular mutants of *Arabidopsis* reveals a role of SERRATE as a mediator. *PLoS Genet.* 5, e703.
- W.D. Ratnasooriya, T.S.P. Fernando, and P.P. Madubashini (2008).** In vitro thrombolytic activity of Sri Lankan black tea, *Camellia sinensis* (L.) O. Kuntze, *J. Nat. Sci. Found. Sri Lanka* 36: 179-181.
- W.H. Briggs, J.D. Folts, and H.E. Osman (2001).** Administration of raw onion inhibits platelet-mediated thrombosis in dogs, *J. Nutr.* 131: 2619-2622.
- Watanabe, M., Tanaka, H., Watanabe, D., Machida, C. and Machida, Y. (2004).** The ACR4 receptor-like kinase is required for surface formation of epidermis-related tissues in *Arabidopsis thaliana*. *Plant J.* 39, 298-308.
- Winter, D., Vinegar, B., Nahal, H., Ammar, R., Wilson, G.V., and Provart, N.J. (2007).** An "Electronic Fluorescent Pictograph" browser for exploring and analyzing largescale biological data sets. *PLoS One.* 2, e718.
- Wu D.H., Shi G.Y., Chuang W.J., Hsu J.M., Young K.C., Chang C.W. (2001).** Coiled coil region of streptokinase gamma-domain is essential for plasminogen activation. *J Biol Chem*; 276: 15025-15033.
- Yadav, A.S. and Bhatnagar, D. (2010).** Inhibition of iron induced lipid peroxidation and antioxidant activity of Indian spices and Acacia in vitro. *Plant foods Hum. Nutr. Mar.* 65: 18 – 24.
- Yamamoto J, Yamada K, Naemura A, Yamashita T, Arai R (2005).** Testing various herbs for antithrombotic effect. *Nutrition*; 21(5): 580-587.
- Yamamoto J, Yamada K, Naemura A, Yamashita T, Arai R. (2005).** Testing various herbs for antithrombotic effect. *Nutrition*, 21(5):580-587.
- Yoshihrio, M., Toshihiro, I., Minpei, K. and Yutaka, S. (1996).** Steroidal Saponins from *Sansevieria trifasciata*. *Phytochemistry* 43: 1325-1331.

- You, M.P., Yang, H.A., Sivasithamparam, K. and Barbetti, M.J. (2009).** A new leaf blight disease of *Trifolium dasyurum* caused by *Botrytis fabae*. *Eur. J. Plant Pathol.* 123, 99-103.
- Yuri J.A., Neira A., Quilodran A., Motomura Y., Palomo I. (2009).** Antioxidant activity and total phenolics concentration in apple peel and flesh is determined by cultivar and agroclimatic growing regions in Chile. *J Food Agri Environ*, 7:3-4.
- Yusuf M, Wahab MA, Yousuf MD, Chowdhury JU, Begum J. (2008).** Some tribal medicinal plants of Chittagong Hill Tracts, Bangladesh. *Bangladesh J Plant Tax*, 14:117-128.
- Zhiguang LI WANGH, Jiazeng L, Guangshen Z, Cunji G. (2000).** Basic and clinical study on the antithrombotic mechanism of glycosaaminoglycan extracted from sea cucumber. *Chin Med J*, 113(8):706-711.
- Zhiguang L.I., WANG Hongli, Jiazeng L.I., Guangshen Z.H.A.N.G., Cunji Gao 6 (2003).** Basic and clinical study on the antithrombotic mechanism of glycosaaminoglycan extracted from sea cucumber. *Chin Med J*; 113(8); 706-711
- Zimmermann, P. Hirsch - Hoffmann, M., Hennig, L. and Gruissem, W. (2004).** Genevestigator Arabidopsis microarray database and analysis toolbox. *Plant Physiol.* 136, 2621-2632.